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Protocol
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## **Hot Phenol RNA Preparation**

## **General Notes**

Ref: Massé, E., Escorcia, F.E., Gottesman, S. (2003) Genes. Dev. 17, 2374-2383.

Always practice RNase-Free technique when working with RNA Use RNA-only pipet tips and tubes Wipe off counter tops and pipetmen with RNaseZap Filter-sterilize solutions prior to use Use either DEPC-H<sub>2</sub>O or MQ-H<sub>2</sub>O Keep RNA cold (4 °C or lower)

Lysis solution (8x) [make ~ 10 mL at a time)] 320 mM NaAcetate (pH 5.0) 8% SDS 16 mM EDTA

## **Protocol:**

- 1. **Pellet** bacteria cells by spinning at 4 °C, 5,000 x g, for 5 minutes.
  - a. If growing bacteria in M9 media, pellet at least 2 mLs.
  - b. If growing bacteria in LB media, you can skip this step and just take 500  $\mu$ L of culture directly to Step 4.
- 2. **Remove** supernatant, being careful not to lose pellet
- 3. **Resuspend** pellet in 500 µL 1x M9
- 4. **Add** 71 μL 8x Lysis Buffer, quickly pipetting up and down to mix. **Immediately add** 570 μL Acid Phenol/Chloroform (Ambion), **vortex**.
- 5. **Incubate** 5 minutes at 65 °C with regular **vortexing** (every 50 sec)
- 6. **Spin** > 12,000 x g, 10 minutes
- 7. **Extract** with phenol/chloroform at least twice until interface is clean

Each time you want to keep the aqueous (top) layer for the next step

Note: the aqueous layer will continually decrease in volume. Add an equal volume of phenol/chloroform each time. E.g. For 500  $\mu L$  aqueous layer, add 500  $\mu L$  phenol/chloroform.

8. After the final extraction, **add** 5 μL glycogen (Ambion) and **precipitate** the RNA in the aqueous layer with 3 volumes of 100% ethanol (-70 °C, 30 minutes)

You can also put the tubes on well-crushed ice in the -20 °C freezer.

- 9. **Spin** 12,000 x g at 4 °C for 10 minutes
- 10. Rinse pellet with 500 µL of 70% ethanol, spin 2 minutes

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- 11. Air dry pellet, with tube inverted on Kimwipe, for 10 minutes
- 12. **Resuspend** pellet in 50 μL DEPC-H<sub>2</sub>O
- 13. Quantitate RNA concentrations

Make a 1:10 dilution in  $H_2O$  and measure absorbance at 260 and 280 nm ( $OD_{260}$  of 1 = 40  $\mu$ g/mL; 260/280 ratio should be between 1.8 and 2.0)